

Gain of Toxicity from ALS/FTD-Linked Repeat Expansions in *C9ORF72* Is Alleviated by Antisense Oligonucleotides Targeting GGGGCC-Containing RNAs

Highlights

- *C9ORF72* repeat expansions cause age-, repeat-size-, and expression-dependent toxicity
- Acquired toxicity, not loss of function, is a major contributor to *C9orf72* disease
- Absence of *C9ORF72* in mice produces splenomegaly and enlarged cervical lymph nodes
- ASO-induced decreases in repeat RNA mitigate *C9ORF72*-associated phenotypes in vivo

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In Brief

Hexanucleotide expansions in *C9ORF72* are the most frequent genetic cause of ALS and FTD. Jiang et al. establish gain of toxicity from repeat-containing RNA, and not loss of *C9ORF72* function, as a central disease mechanism and establish the feasibility of ASO-mediated therapy.



Gain of Toxicity from ALS/FTD-Linked Repeat Expansions in *C9ORF72* Is Alleviated by Antisense Oligonucleotides Targeting GGGGCC-Containing RNAs

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SUMMARY

Hexanucleotide expansions in *C9ORF72* are the most frequent genetic cause of amyotrophic lateral sclerosis and frontotemporal dementia. Disease mechanisms were evaluated in mice expressing *C9ORF72* RNAs with up to 450 GGGGCC repeats or with one or both *C9orf72* alleles inactivated. Chronic 50% reduction of *C9ORF72* did not provoke disease, while its absence produced splenomegaly, enlarged lymph nodes, and mild social interaction deficits, but not motor dysfunction. Hexanucleotide expansions caused age-, repeat-length-, and expression-level-dependent accumulation of RNA foci and dipeptide-repeat proteins synthesized by AUG-independent translation, accompanied by loss of hippocampal neurons, increased anxiety, and impaired cognitive function. Single-dose injection of antisense

oligonucleotides (ASOs) that target repeat-containing RNAs but preserve levels of mRNAs encoding *C9ORF72* produced sustained reductions in RNA foci and dipeptide-repeat proteins, and ameliorated behavioral deficits. These efforts identify gain of toxicity as a central disease mechanism caused by repeat-expanded *C9ORF72* and establish the feasibility of ASO-mediated therapy.

INTRODUCTION

Amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) are two devastating adult-onset neurodegenerative diseases with distinct clinical features but common pathological features and genetic causes (Ling et al., 2013). Hexanucleotide GGGGCC repeat expansions in a noncoding region of the *C9ORF72* gene are the most common inherited cause of ALS and FTD (DeJesus-Hernandez et al., 2011; Renton et al., 2011).

Proposed mechanisms by which *C9ORF72* repeat expansions cause disease (referred to as C9ALS/FTD) include loss of C9ORF72 protein function and gain of toxicity (Gendron et al., 2014; Ling et al., 2013).

A reduction in C9ORF72 function (i.e., haploinsufficiency) is supported by decreased expression of *C9ORF72* mRNAs in patient tissues resulting from DNA and histone hypermethylation with partial silencing of the repeat-containing allele (Belzil et al., 2013; DeJesus-Hernandez et al., 2011; Gijssels et al., 2012; Liu et al., 2014; Xi et al., 2013, 2015). RNA gain of toxicity has been proposed to arise from folding of repeat-containing RNAs into stable structures that sequester RNA-binding proteins to nuclear RNA foci, a mechanism originally established for myotonic dystrophy (reviewed by Wojciechowska and Krzyzosiak, 2011). Indeed, foci containing sense GGGGCC or antisense GGCCCC repeat RNA are present in tissues from C9ALS/FTD patients (DeJesus-Hernandez et al., 2011; Gendron et al., 2013; Lagier-Tourenne et al., 2013; Mizielińska et al., 2013; Mori et al., 2013a; Zu et al., 2013). Several RNA-binding proteins appear enriched in sense RNA foci (Cooper-Knock et al., 2014; Donnelly et al., 2013; Lee et al., 2013; Mori et al., 2013b; Sareen et al., 2013; Xu et al., 2013), but evidence supporting a loss of function of these proteins is lacking.

Another proposed disease mechanism from *C9ORF72* repeat expansions is the production of aberrant dipeptide-repeat (DPR) proteins through repeat-associated non-AUG-dependent (RAN) translation, a phenomenon discovered in spinocerebellar ataxia type 8 and myotonic dystrophy (Zu et al., 2011). In C9ALS/FTD patients, five DPR proteins—poly(GA), poly(GR), poly(GP), poly(PA), and poly(PR)—are produced from all reading frames of either sense or antisense repeat-containing RNAs (Ash et al., 2013; Gendron et al., 2013; Mori et al., 2013a, 2013c; Zu et al., 2013). DPR proteins form p62-positive, TDP-43-negative inclusions, with poly(GA), poly(GP), and poly(GR) aggregates being the most abundant (Mackenzie et al., 2014; Mori et al., 2013c). Poly(GP) and poly(GA) can also be detected by immunassay in patient-derived cultured cells, postmortem tissues, and cerebrospinal fluid (Gendron et al., 2015; Su et al., 2014; van Blitterswijk et al., 2015). Overexpressing (Mizielińska et al., 2014; Wen et al., 2014; Yang et al., 2015) or exogenously exposing (Kwon et al., 2014) cells and other model systems to high levels of arginine-containing DPR proteins induces rapid cell death. Other studies have reported poly(GA) to be particularly prone to aggregation and to confer toxicity (Gendron et al., 2015; May et al., 2014; Schludi et al., 2015; Zhang et al., 2014, 2016). Lastly, a recent flurry of studies found that nucleocytoplasmic transport is impaired in C9ALS/FTD (Boeynaems et al., 2016; Freibaum et al., 2015; Jovičić et al., 2015; Zhang et al., 2015, 2016), although the exact roles of RNA foci and/or DPR proteins in this pathway are not yet resolved. Indeed, divergent outcomes were reported regarding the toxicity of repeat-containing RNAs or DPR proteins in cell culture (Devlin et al., 2015; Kwon et al., 2014; Lee et al., 2013; May et al., 2014; Rossi et al., 2015; Schludi et al., 2015; Wen et al., 2014; Zhang et al., 2014; Zu et al., 2013), flies (Freibaum et al., 2015; Mizielińska et al., 2014; Tran et al., 2015; Xu et al., 2013; Zhang et al., 2015), yeast (Jovičić et al., 2015), and mice (Chew et al., 2015; Hukema et al., 2014; O'Rourke et al., 2015; Peters et al., 2015; Zhang et al., 2016).

By production and analysis of mice in which one or both *C9orf72* alleles were inactivated, and multiple transgenic mouse lines expressing up to 450 *C9ORF72* hexanucleotide repeats, we tested disease mechanism(s) associated with C9ALS/FTD. Herein we show a repeat-length-dependent gain of toxicity, while chronic reduction of C9ORF72 does not produce nervous system disease. Single-dose injection of antisense oligonucleotides (ASOs) that selectively target repeat-containing RNAs causes a rapid reduction in RNA foci and DPR proteins, while maintaining overall levels of C9ORF72-encoding mRNAs and producing a sustained alleviation of behavioral deficits.

RESULTS

Reduction in C9ORF72 Is Well-Tolerated, but Splenomegaly and Enlarged Lymph Nodes Develop in Its Absence

To determine the consequence of loss of C9ORF72 function in vivo, mice were generated in which a LacZ reporter replaced exons 2–6 of the endogenous *C9orf72* gene (Figure 1A). As expected, total *C9orf72* RNAs and the full-length 54 kD C9ORF72 protein were reduced to 50% in brains of heterozygous *C9orf72*^{+/-} mice and were completely absent in homozygous *C9orf72*^{-/-} mice (Figures 1B and 1C). A β -galactosidase activity assay, used to assess endogenous *C9orf72* gene expression throughout the CNS, revealed broad expression of *C9orf72* not only in neurons of the gray matter as previously reported (Suzuki et al., 2013), but also in glial cells of white matter in various CNS regions, including the cerebellum and spinal cord (Figures S1A and S1B, available online). Further evidence confirming substantial expression of the *C9orf72* gene in glial cells was obtained using high-throughput sequencing of translated RNAs purified from BacTRAP transgenic mice that express EGFP-tagged ribosomal protein L10a in defined cell populations (Heiman et al., 2008). *C9orf72* RNAs were identified in all three cell types tested, with relative abundances of 2.5:1:1 in motor neurons, astrocytes, and oligodendrocytes, respectively (Figure S1C). No overt neuropathology developed in *C9orf72*^{+/-} or *C9orf72*^{-/-} mice, and no increase in GFAP-positive astrocytes or IBA-1-positive microglia occurred in brain and spinal cord (Figures S1D and S1E).

The reduction in C9ORF72 to 50% of its normal level, as reported in C9ALS/FTD patients, was well tolerated with *C9orf72*^{+/-} mice surviving into adulthood with a normal body weight and no signs of disease (Figures 1D–1F). In contrast, although *C9orf72*^{-/-} mice were born in the expected Mendelian ratio with no change in survival through 11 months of age, only 7% of mice survived to 20 months of age (Figure 1D). *C9orf72* null mice showed reduced body weight (Figure 1E), splenomegaly (Figure 1F), and enlarged cervical lymph nodes (Figure S1F) by 12 months of age. The spleen size was comparable in *C9orf72*^{+/-} mice (80.33 \pm 0.88 mg, n = 3, mean \pm SEM) versus wild-type littermates (80 \pm 0.41 mg, n = 4) at 18 months of age. Staining of CD3-positive T lymphocytes and CD45R/B220-positive B lymphocytes revealed disrupted architectures of the spleen and cervical lymph nodes in *C9orf72*^{-/-} mice (Figures S1G and S1H). Despite the enlarged size, the total number of lymphoid cells was not changed in the spleen, while the number

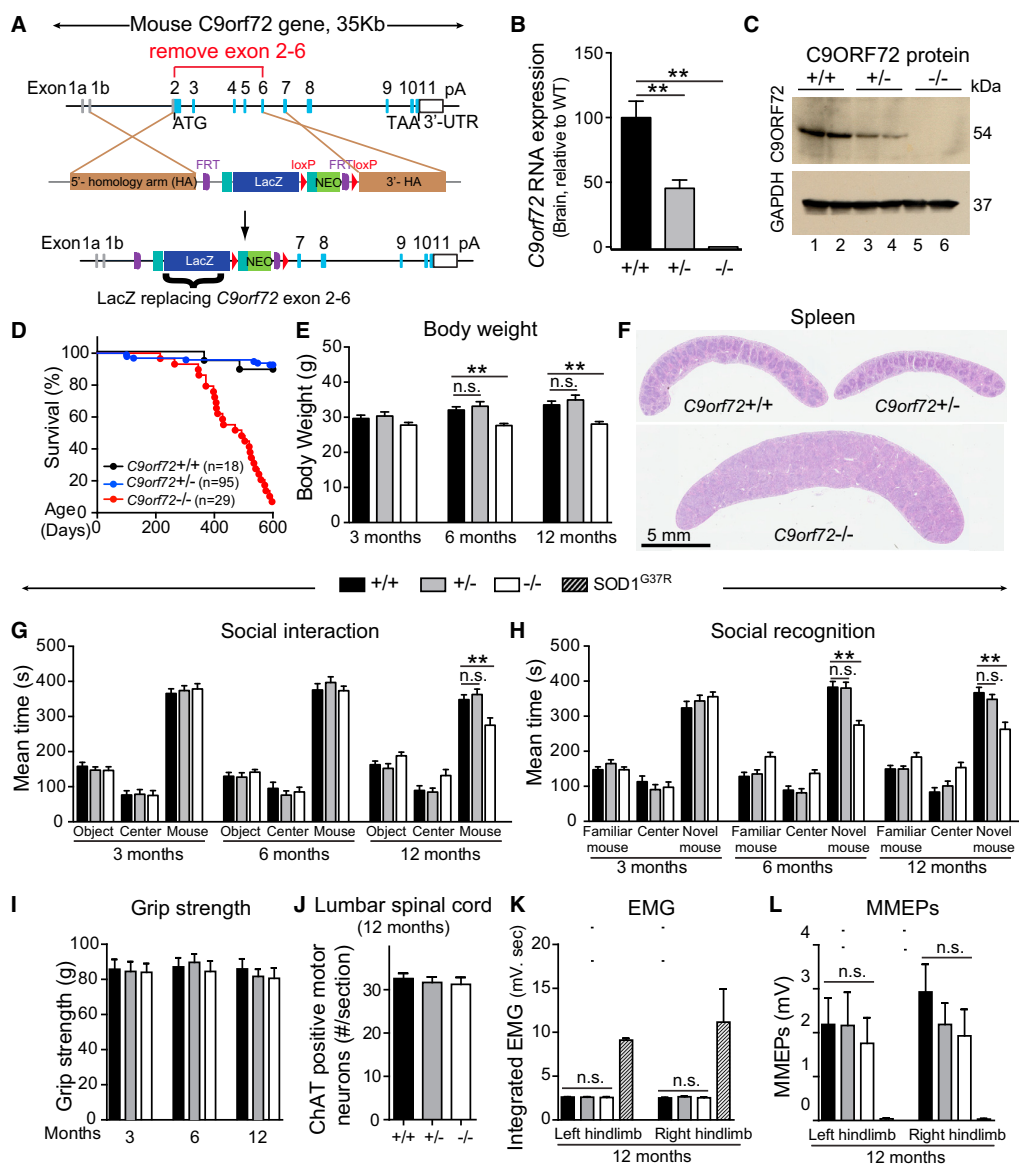


Figure 1. Reduction of C9ORF72 Is Well-Tolerated, but Complete Loss of C9ORF72 Induces Premature Death and Splenomegaly

(A) Targeting strategy to generate *C9orf72* null mice with exons 2–6 replaced with β -galactosidase and neomycin (Neo) genes. Schematic shows (upper panel) the genomic region, (middle panel) targeting construct, and (lower panel) final targeted *C9orf72* gene. Filled boxes correspond to exons. Open box represents the 3' untranslated region (3'-UTR).

(B) qRT-PCR measurement of mouse *C9orf72* RNAs in brains of *C9orf72*^{+/+}, *C9orf72*^{+/-}, and *C9orf72*^{-/-} mice (n = 6 per group).

(C) Immunoblot demonstrating the levels of the 54 kD mouse C9ORF72 protein. GAPDH was used as a loading control.

(D) Survival curve up to 600 days.

(E) Body weights at 3, 6, and 12 months of age (n = 25 per genotype).

(F) Spleen sizes at 12 months of age.

(G–I) Behavioral performance measured at 3, 6, and 12 months of age (n = 25 animals per group). (G) Social interactions measured by the mean time spent with an object, in the center, or with a mouse. (H) Social recognition measured by the mean time spent with a familiar mouse, in the center, or with a novel mouse.

(I) Hindlimb grip strength.

(J) Average number of ChAT-positive neurons per section in the anterior horn of lumbar spinal cord in 12-month-old mice (n = 4–5 per group).

(K and L) Resting electromyographic (EMG) recordings (K) and myogenic motor-evoked potentials (MMEPs) (L) in 12-month-old *C9orf72* mice (n = 5 per genotype) or in end-stage transgenic mice expressing mutant SOD1^{G37R} (crosshatched bars, n = 2). Error bars represent SEM in biological replicates; n.s., not significant; *p < 0.05, **p < 0.01 using one-way ANOVA.

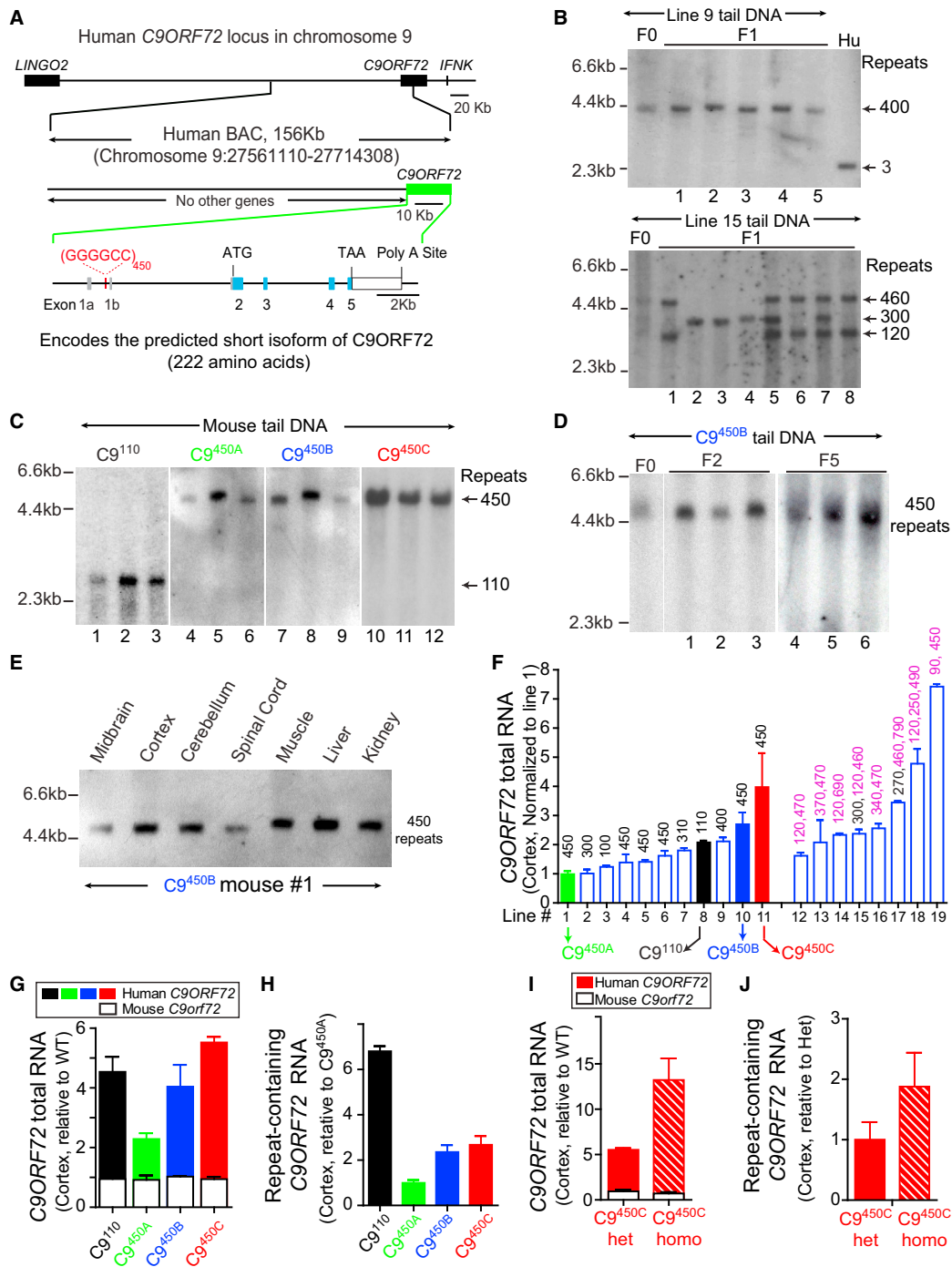


Figure 2. Generation of Multiple BAC Transgenic Mouse Lines Expressing Different Levels of a Human *C9ORF72* Transgene with 100–700 GGGGCC Repeats

(A) Schematic of the human BAC containing 450 GGGGCC repeats in the first intron of a truncated human *C9ORF72* gene. The coordinates of the BAC sequence on the University of California, Santa Cruz Genome Browser (Hg19) are indicated. No other gene is on the BAC.

(legend continued on next page)

of myeloid cells increased dramatically. Both lymphoid and myeloid cells were increased by more than an order of magnitude in the cervical lymph nodes (Figure S2A). In addition, *C9orf72* null mice showed decreased hemoglobin and packed cell volume, decreased percentage of lymphocytes, and increased percentage of neutrophils in blood (Figure S2B).

To determine whether partial or complete loss of *C9ORF72* triggered age-dependent cognitive and/or motor deficits, longitudinal assessment of strength, motor coordination, anxiety, sociability, and learning functions in cohorts of *C9orf72*^{+/-}, *C9orf72*^{2+/-}, and *C9orf72*^{-/-} mice was performed. No behavioral abnormalities were observed in *C9orf72*^{+/-} mice at any of the ages tested (3, 6, and 12 months) (Figures 1G–1I and S2C–S2H). By 12 months of age, *C9orf72* null mice developed mild social interaction (Figure 1G) and social recognition (Figure 1H) abnormalities compared with wild-type or heterozygous littermates. In addition, 12-month-old *C9orf72*^{-/-} mice developed mild motor deficits on a rotarod assay (Figure S2C) without differences in grip strength, gait, or general activity (Figures 1I, S2D, and S2E), or loss of ChAT-positive lower motor neurons (Figure 1J). Resting electromyographic (EMG) recordings were also normal in the gastrocnemius muscles of mice lacking *C9ORF72*, consistent with intact neuromuscular connectivity (Figure 1K). The amplitudes of myogenic motor-evoked potentials (MMEPs), a measure of connectivity of the entire neuromuscular unit, are comparable among *C9orf72*^{+/-}, *C9orf72*^{-/-}, and *C9orf72*^{+/+} mice (Figure 1L), in contrast to their almost complete loss in end-stage *SOD1*^{G37R} mice. Overall, *C9ORF72* reduction alone is not sufficient to cause C9ALS/FTD-associated phenotypes in mice.

BAC *C9ORF72* Transgenic Mouse Lines with Different Repeat Sizes and Expression Levels

To test a potential gain of toxicity from repeat expansions, transgenic mice were generated that express a bacterial artificial chromosome (BAC) with the human expanded *C9ORF72* gene from a C9ALS patient. The BAC includes 140 kb of sequence 5' to the *C9ORF72* exon 1a (including what is likely to be the complete promoter region) and exons 1–5 of the gene. The BAC does not encode the 54 kD *C9ORF72* protein (Figure 2A) (DeJesus-Hernandez et al., 2011). The interferon kappa gene, which lies within 23 kb 3' of *C9ORF72* on human chromosome 9, is not present, nor is any other known gene besides *C9ORF72*.

Thirty-two founder mice were generated in a C57BL6/C3H hybrid background and backcrossed to C57BL/6 mice. Founders from eight lines had either multiple repeat sizes that separated in subsequent generations (Figure 2B) or multiple

transgene copies with different repeat lengths at the same locus that segregated together (Figure S3A). Eleven lines contained a single repeat size that was stably inherited from filial (F0) to F5 generations in the large majority of mice (Figures 2B–2D), albeit analysis of 42 littermates from one line identified three repeat contraction events (Figure S3B). At most, modest expansion was found within the CNS or peripheral tissues (Figures 2E, S3C, and S3D), in contrast to humans, for whom somatic heterogeneity and repeat instability have been reported, especially within the CNS (van Blitterswijk et al., 2013).

Transgene expression levels were examined in 19 lines carrying between 110 and 790 repeats (Figure 2F). Recognizing that transgenes with multiple repeat sizes would preclude correlation analyses of repeat-length- and expression-associated toxicity, we selected four lines with defined repeat lengths and/or different transgene expression levels: line 8 with ~110 repeats (hereafter designated C9¹¹⁰) and three lines expressing various levels of RNAs but each containing ~450 repeats (lines 1, 10, and 11, designated C9^{450A}, C9^{450B}, and C9^{450C}, respectively). C9¹¹⁰ and C9^{450B} have comparable levels of transgene expression but different repeat sizes, whereas C9^{450B} has a similar repeat size as C9^{450A}, but higher transgene expression (Figures 2C and 2F). *C9ORF72* transgene-encoded RNAs were three to four times the level of endogenous mouse *C9orf72* RNA in lines C9¹¹⁰, C9^{450B}, and C9^{450C}, and equal to endogenous *C9orf72* in line C9^{450A} (Figure 2G). Endogenous *C9orf72* RNAs were unchanged in all lines. Similar to total *C9ORF72* RNAs, repeat-containing RNAs (measured by qRT-PCR with variant-specific primers; Figure S3E) were higher in C9^{450B} mice than C9^{450A} mice. Repeat-containing RNAs were three times higher in C9¹¹⁰ than C9^{450B} mice (Figure 2H), despite comparable total *C9ORF72* RNA levels (Figures 2G and S3F). Breeding produced homozygous C9^{450C} mice with *C9ORF72* expression ~12 times the level of mouse *C9orf72* (Figures 2I and 2J).

Repeat Size- and Dose-Dependent Accumulation of Sense and Antisense RNA Foci

RNAs transcribed from both sense (GGGGCC) and antisense (GGCCCC) strands of *C9ORF72* have been reported to accumulate into RNA foci in cultured cells and postmortem tissues from C9ALS/FTD patients (Gendron et al., 2013; Lagier-Tourenne et al., 2013; Mizielska et al., 2013; Zu et al., 2013). Use of fluorescence in situ hybridization detected sense and antisense foci in all three mouse lines expressing 450 repeats. Foci were found throughout the CNS, including the frontal cortex, hippocampus, cerebellum, and spinal cord, as early as 2 months of age (Figure 3A) and were not observed in wild-type mice (Figure S3G).

(B–D) Genomic DNA blot analysis of tail DNA from (B) founder (F0) and F1 transgenic mice from lines 9 or 15 and DNA from human fibroblasts (Hu) with normal *C9ORF72* alleles; (C) twelve different mice of lines 1, 8, 10, and 11 (redesignated C9^{450A}, C9¹¹⁰, C9^{450B}, and C9^{450C}, respectively); and (D) mice from F0, F2, and F5 generations in line C9^{450B}.

(E) Repeat lengths determined by genomic DNA blotting using DNA from the CNS and peripheral tissues of a C9^{450B} mouse.

(F) Human *C9ORF72* RNA in cortex of transgenic mice measured by qRT-PCR, normalized to C9^{450A} mice. Numbers above bars are repeat lengths measured by genomic DNA blots. Numbers in pink represent transgenes with different repeat lengths at the same locus that segregated together.

(G) Expression levels of total (human plus mouse) *C9ORF72* RNAs in the cortex of C9¹¹⁰, C9^{450A}, C9^{450B}, and C9^{450C} mice normalized to the level of endogenous *C9orf72* RNA.

(H) Level of repeat-containing *C9ORF72* RNA variants in the cortex measured by qRT-PCR and normalized to levels in C9^{450A} mice.

(I and J) Levels of (I) total *C9ORF72* RNAs (human plus mouse) normalized to *C9orf72* levels in wild-type littermates, or (J) repeat-containing *C9ORF72* RNA measured by qRT-PCR in the cortex of heterozygous and homozygous C9^{450C} mice. Error bars represent SEM from 3 to 5 biological replicates per group.

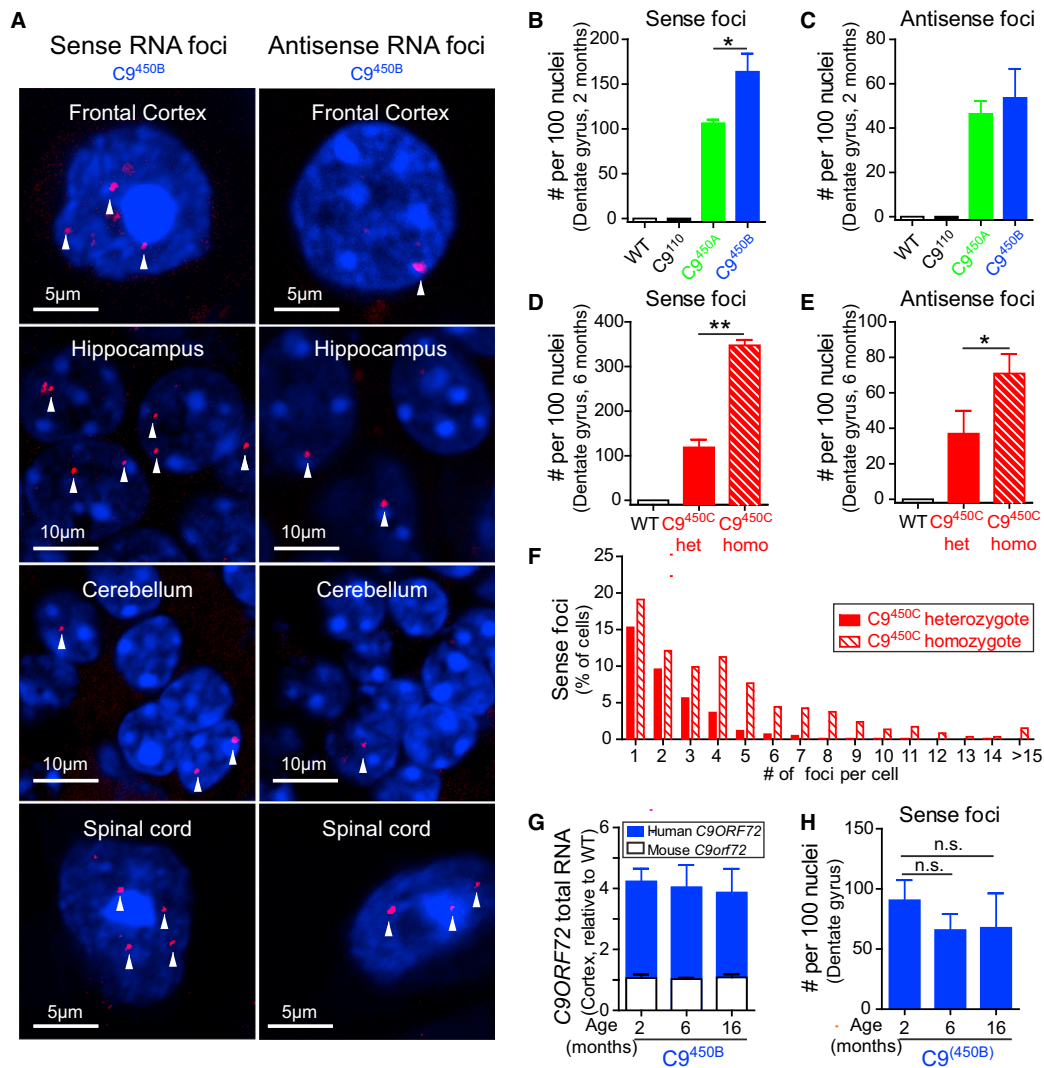


Figure 3. Repeat Size- and Dose-Dependent Accumulation of Sense and Antisense RNA Foci in *C9ORF72* Transgenic Mice

(A) FISH detection of sense and antisense RNA foci (arrows) in 2-month-old *C9^{450B}* mice. DNA was stained with DAPI.

(B–E) Numbers of sense and antisense foci (per 100 nuclei) in hippocampal dentate gyrus of (B and C) 2-month-old *C9¹¹⁰*, *C9^{450A}*, and *C9^{450B}* mice, and (D and E) 6-month-old heterozygous and homozygous *C9^{450C}* mice. Error bars represent SEM in 3–5 biological replicates. * $p < 0.05$, ** $p < 0.01$ using Student's *t* test.

(F) Quantification of sense RNA foci per nucleus in hippocampal dentate gyrus of 6-month-old heterozygous and homozygous *C9^{450C}* mice.

(G) qRT-PCR measurement of human and mouse *C9ORF72* RNAs in cortex of 2-, 6-, and 16-month-old *C9^{450B}* mice, normalized to wild-type littermates ($n = 2–4$ per group).

(H) Numbers of sense foci (per 100 nuclei) in hippocampal dentate gyrus of 2-, 6-, and 16-month-old *C9^{450B}* mice. Error bars represent SEM in 2–4 biological replicates. n.s., not significant using one-way ANOVA.

Sense foci were most abundant in the frontal cortex, followed by the hippocampal dentate gyrus, retrosplenial cortex, and molecular layer of the cerebellum (Figure S3H). An unexpected, surprisingly strong influence of repeat length on foci formation was uncovered: whereas mice with 450 repeats developed foci (Figure 3A), neither sense nor antisense foci were detected in any brain region, or at any age, in *C9¹¹⁰* mice with 110 repeats (Figures 3B and 3C). This was despite the fact that transgene

expression in *C9¹¹⁰* mice was 3.5 times that of endogenous *C9orf72* RNAs (seven times the level of RNA from a single endogenous *C9orf72* allele) (Figure 2G) and that levels of repeat-containing RNAs in *C9¹¹⁰* mice were three and six times higher than in *C9^{450B}* and *C9^{450A}* mice, respectively (Figure 2H).

Doubling the level of 450 repeat-containing sense strand RNAs (compare RNA levels in *C9^{450B}* versus *C9^{450A}*; Figures 2G and 2H) doubled sense foci accordingly (Figure 3B). Similarly,

doubling repeat-containing RNAs by generating homozygous C9^{450C} mice more than doubled the overall number of sense foci (Figure 3D) and the number of foci per cell (Figure 3F). The proportion of cells that developed sense foci also increased from 52% in the dentate gyrus of heterozygous C9^{450C} mice to 81% in homozygous mice (Figure S3I), and some cells accumulated more than 30 sense foci (Figure 3F). Transgene expression and foci burden remained constant with age (Figures 3G and 3H).

Antisense RNAs from the human *C9ORF72* transgene, identified using a strand-specific RT-PCR strategy (Figures S3K and S3L), also accumulated into RNA foci (Figure 3A). As with sense foci, antisense foci increased with increased repeat length (Figure 3C) and RNA expression level, with both the overall number of antisense foci (Figure 3E) and the percentage of cells with foci (Figures S3I and S3J) nearly doubling in homozygous versus heterozygous C9^{450C} mice.

Age-, Repeat-Length-, and Expression-Dependent Cytoplasmic Inclusions of DPR Proteins

Inclusions of DPR proteins produced by RAN translation from GGGGCC or GGCCCC repeats are a neuropathological hallmark of C9ALS/FTD (Ash et al., 2013; Gendron et al., 2013; Mori et al., 2013a, 2013c; Zu et al., 2013). As in human patients, perinuclear, cytoplasmic aggregates of sense strand RNA-encoded poly(GA), poly(GP), or poly(GR) proteins were detected in multiple CNS regions in C9^{450C} mice as young as 3 months of age (Figures 4A and 4B). Poly(GA) and poly(GP) aggregates in 22-month-old C9^{450C} mice were most abundant in the retrosplenial cortex, followed by the hippocampal dentate gyrus and frontal cortex (Figures 4C and S4A). Fewer poly(GA) aggregates were observed in the cerebellum (Figure S4B) or spinal cord. Dot blot assay confirmed accumulation of poly(GP) in cortical lysates from 6-month-old C9^{450C} mice (Figure S4C). As in C9ALS/FTD patients (Mori et al., 2013a), DPR proteins coaggregated into common, p62-containing inclusions (Figures 4D, 4E, and S4D). Antisense strand-encoded poly(PR) and poly(PA) proteins have only been detected in rare aggregates in patient samples (Gendron et al., 2013; Mackenzie et al., 2015; Mori et al., 2013a; Schludi et al., 2015; Zu et al., 2013). None were detected in C9 mice of any line or at any age.

Similar to RNA foci formation, DPR protein expression was dependent on repeat length, as established by immunoassay for quantitative measurements of poly(GP) (Su et al., 2014). Although C9¹¹⁰ mice expressed a 3-fold higher level of repeat-containing *C9ORF72* RNAs than C9^{450B} mice (Figure 2H), no poly(GP) was detected in 2% SDS-soluble homogenates prepared from tissues of various neuroanatomical regions from C9¹¹⁰ mice, while poly(GP) was easily detected in mice expressing 450 repeats (Figure 4F). As expected, more poly(GP) was detected in mice with higher *C9ORF72* RNA expression (compare C9^{450A} and C9^{450B} mice; Figure 4F; and homozygous C9^{450C} versus age-matched heterozygotes; Figure 4G).

SDS-soluble poly(GP) decreased with age in C9^{450A} and C9^{450B} mice (Figure 4H), despite constant transgene expression (Figure 3G), indicating that DPR proteins may become insoluble over time. Consistent with this notion, poly(GA) aggregated more in the retrosplenial cortex (Figure 4I) and hippocampus (Fig-

ure S4E) as mice aged, and the size of poly(GA) inclusions increased over time (Figure 4J). Age-dependent increases in the number and size of poly(GP) and poly(GR) aggregates were also detected (Figures S4F and S4G). In addition, more cells accumulated DPR protein aggregates in 3-month-old C9^{450C} homozygous mice compared to heterozygous mice, and the abundance of such aggregates was increased at 6 months of age (Figures 4I, 4K, S4F, and S4G).

Age-Dependent Cognitive Impairment in C9 Mice Expressing 450 Repeats

C9 transgenic mice were tested for age-dependent disease. No significant motor deficits or weight loss were observed by 18 months of age in two mouse lines expressing 450 repeats (C9^{450B} or C9^{450C}; Figures 5A–5C, S5A, and S5B). By 12 months of age, no loss of ChAT-positive lower motor neurons (Figures 5D and 5E) or increased glial activation (Figure S5C) was detected in the spinal cord of C9^{450C} mice, which express the highest levels of transgene. Functional innervation and connectivity of the entire neuromuscular unit were normal, as measured by resting EMG recordings and MMEP amplitudes, respectively, in 12-month-old C9¹¹⁰ and C9^{450C} mice (Figures S5D and S5E). There was no loss of CTIP-2-positive upper motor neurons in layer 5 of the motor cortex in either line (Figure S5F).

Cognitive and behavioral dysfunction, however, developed in an age-dependent manner. While 4-month-old C9^{450B} mice performed as well as wild-type littermates in an assay that measures spatial learning and memory (the Barnes maze), both C9^{450B} and C9^{450C} lines developed deficits by 12 months of age that were sustained to 18 months (Figure 6A). A second spatial-learning assay (the radial arm maze) confirmed a working memory deficit at 12 and 18 months of age in C9^{450B} and C9^{450C} mice (Figure 6B). Aged C9^{450B} and C9^{450C} male mice also displayed abnormalities in two “anxiety” assays, a marble burying test (Figure 6C) and an elevated plus maze (Figure 6D). The spatial learning and anxiety abnormalities were not accompanied by behavioral differences in social interaction, social recognition, or social communication, nor did assays of novel object recognition, fear conditioning, and serial reversal learning uncover any impairment (Figures S6C–S6H).

Recognizing the crucial role of the hippocampus in spatial learning and memory (Sharma et al., 2010), we tested the C9 mice for age-dependent hippocampal neuron loss. Blinded quantification in two hippocampal regions (the dentate gyrus and CA1 region) of 4- and 12-month-old C9^{450B} and C9^{450C} mice identified mild, age-dependent neuronal loss (Figures 6E and 6F). Neuronal loss was repeat-length dependent, as no loss was seen in C9¹¹⁰ mice that expressed higher levels of repeat-containing RNAs than C9^{450C} mice (Figure S7A). No astrogliosis or microgliosis was observed, and neuronal loss was region specific, as the neuronal density in the retrosplenial cortex remained unchanged (Figure S7A). Although levels of both sarkosyl-soluble and sarkosyl-insoluble (but urea-soluble) phosphorylated TDP-43 (pTDP-43) were significantly increased in 22-month-old C9^{450C} mice (Figure S7C), no TDP-43 mislocalization or aggregation was observed (Figure S7B). Similarly, no mislocalized, discontinuous, or punctate aggregates of RanGAP1 or Lamin B (Figures S7D and S7E) were identified.

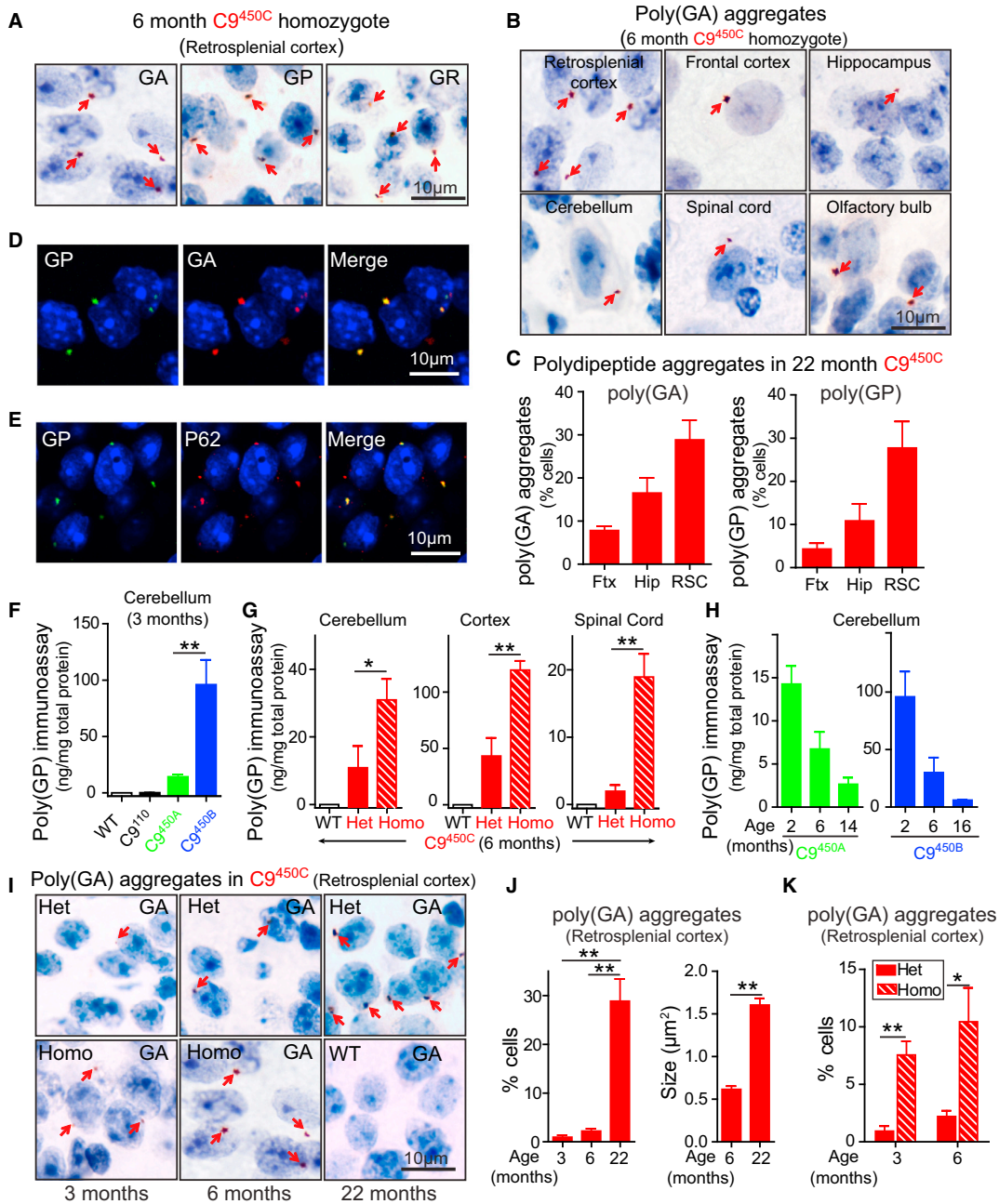


Figure 4. Repeat Size- and Expression-Dependent Production of Sense-Strand-Encoded DPR Proteins Is Associated with Age-Dependent Formation of Cytoplasmic Inclusions

(A) Poly(GA), poly(GP), and poly(GR) perinuclear aggregates (arrows) detected by immunohistochemistry in the retrosplenial cortex of 6-month-old homozygous **C9^{450C}** mice. Nuclei were stained with haemalum.
 (B) Poly(GA) aggregates (arrows) in different CNS regions of 6-month-old homozygous **C9^{450C}** mice.
 (C) Percent of cells containing poly(GA) or poly(GP) inclusions in frontal cortex (Ftx), hippocampus (Hip), and retrosplenial cortex (RSC) of 22-month-old heterozygous **C9^{450C}** mice (n = 2–4 biological replicates).
 (D and E) (D) Aggregates positive for (green) poly(GP), (red) poly(GA), or (yellow) both, and (E) (green) poly(GP) and (red) P62-positive inclusions identified by immunofluorescence in retrosplenial cortex of 6-month-old homozygous **C9^{450C}** mice. DNA is stained with DAPI.

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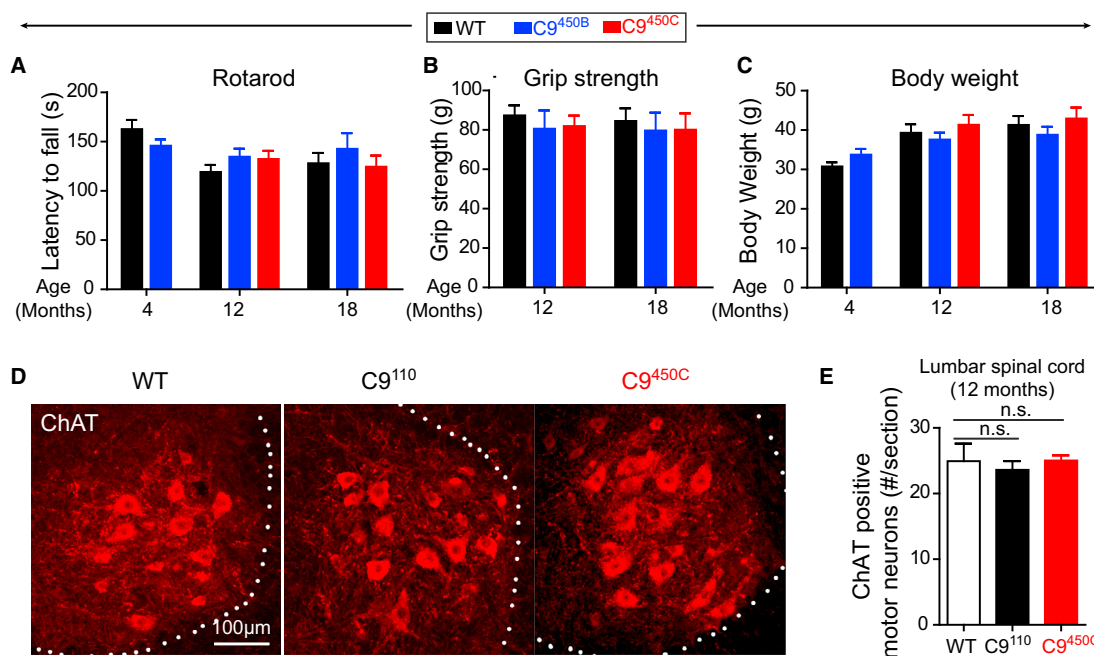


Figure 5. No Motor Neuron Loss and Motor Deficits in *C9ORF72* Transgenic Mice with 450 Repeats

(A–C) (A) Motor performance on a rotarod measured by latency to fall, (B) hindlimb grip strength, and (C) body weight of 4-, 12-, and 18-month-old wild-type (WT), C9^{450B}, and C9^{450C} mice.

(D) Choline acetyltransferase (ChAT)-positive motor neurons detected by immunofluorescence in the anterior horn of lumbar spinal cord of 12-month-old WT, C9¹¹⁰, and C9^{450C} mice.

(E) Average number of ChAT-positive neurons per section. Error bars represent SEM in $n = 4–5$ animals per group. n.s., not significant using one-way ANOVA.

Single-Dose ASOs Reduce Foci, DPR Proteins, and Behavioral Deficits in C9 Mice

ASOs mediate cleavage of target RNAs through action of the primarily nuclear enzyme RNase H, and ASO therapy has gone to clinical trial for ALS (Miller et al., 2013; Smith et al., 2006) and at least two neurodegenerative disorders linked to repeat expansions: myotonic dystrophy (Wheeler et al., 2012) and Huntington's disease (Kordasiewicz et al., 2012). To test whether in vivo administration of ASOs can mediate RNase H-dependent, selective reduction of *C9ORF72* hexanucleotide repeat-containing RNAs in the rodent CNS, 3-month-old C9^{450B} mice were treated with a single intraventricular bolus injection of ASOs targeting human *C9ORF72* RNAs or a control ASO that does not have any target in the mouse genome (Figure 7A). One *C9ORF72* ASO (ASO1) was complementary to a sequence that partially overlaps the 5' end of the hexanucleotide expansion and specifically targets repeat-containing *C9ORF72* RNA variants. A sec-

ond ASO (ASO2) hybridizes within exon 2 and thus targets all *C9ORF72* RNA variants (Figure 7B). Four weeks after a single injection, both ASO1 and ASO2 decreased repeat-containing *C9ORF72* RNA levels in the cortex and spinal cord to 20%–40% of levels in control ASO-treated mice (Figure 7C). As expected, ASO2 decreased total and exon 1b-containing *C9ORF72* RNAs by 40%–60%. However, ASO1, which targets repeat RNA, preserved exon 1b-containing, *C9ORF72* protein-encoding RNAs (Figure 7E) and 80% of total *C9ORF72* RNAs (Figure 7D).

Within 4 weeks after a single injection of either ASO1 or ASO2, the number of sense foci was reduced to 40%–45% (Figures 7F and 7G). Antisense RNA foci remained unchanged (Figure 7F). Both SDS-soluble poly(GP) and poly(GA) were decreased to almost undetectable levels in the cortex and spinal cord (Figures 7H and 7I). The profound reduction of poly(GP), which can be translated from sense or antisense strand RNAs, demonstrates

(F–H) Levels of poly(GP) soluble in 2% SDS measured by immunoassay in (F) cerebellum of 3-month-old C9¹¹⁰, C9^{450A}, and C9^{450B} mice; (G) cerebellum, cortex, and spinal cord of 6-month-old heterozygous and homozygous C9^{450C} mice; and (H) during aging in the cerebellum of C9^{450A} and C9^{450B} mice ($n = 2–5$ biological replicates).

(I) Poly(GA) aggregates (arrows) identified by immunohistochemistry in retrosplenial cortex of heterozygous or homozygous C9^{450C} mice or wild-type mice at the noted ages.

(J) (Left panel) Percent of cells containing poly(GA) aggregates and (right panel) average size of poly(GA) inclusions in retrosplenial cortex of C9^{450C} mice ($n = 2–3$ biological replicates).

(K) Percent of cells with poly(GA) inclusions in retrosplenial cortex of heterozygous and homozygous C9^{450C} mice ($n = 3$ per group). Error bars represent SEM. * $p < 0.05$, ** $p < 0.01$ using Student's t test.

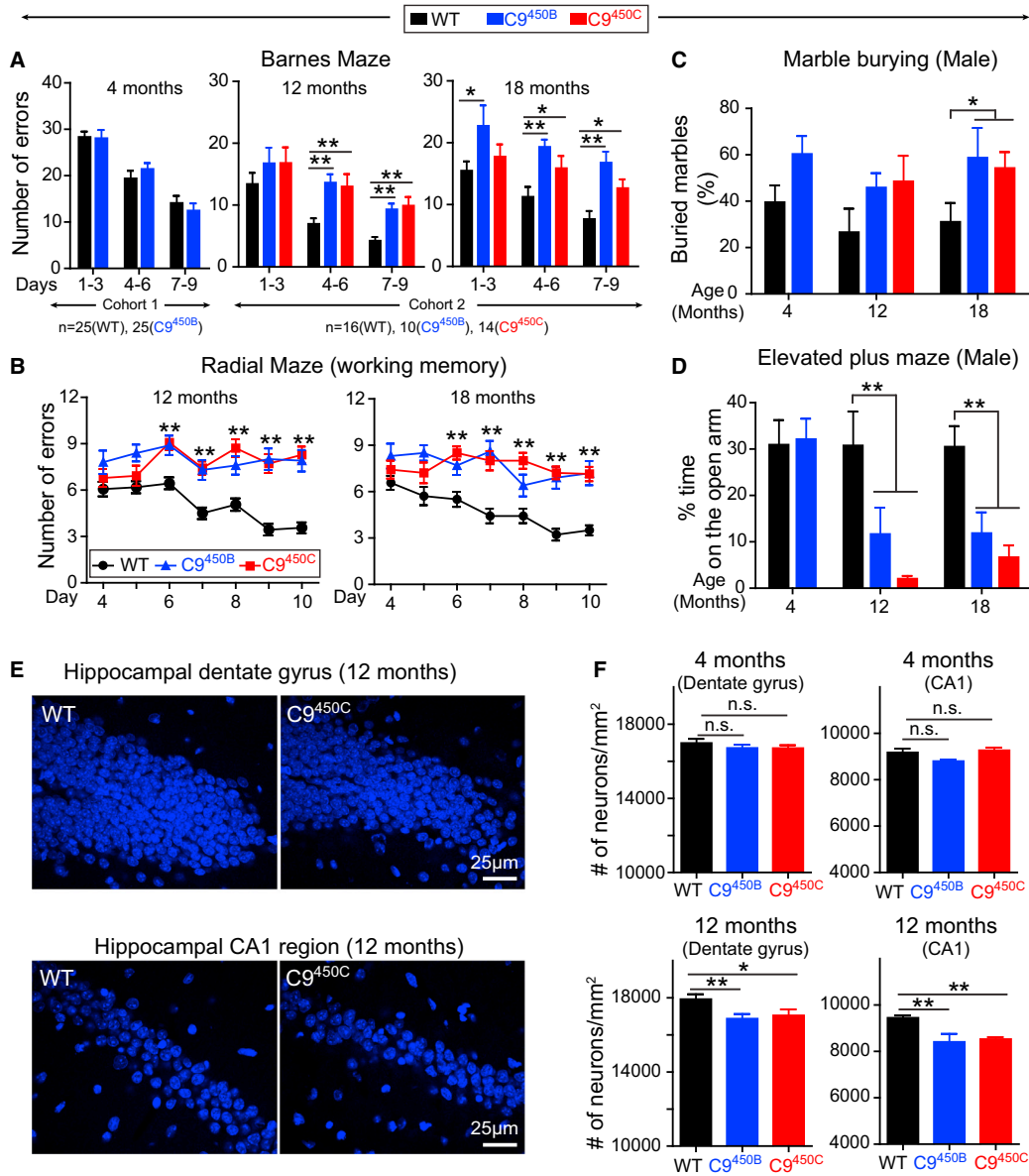


Figure 6. Age-Dependent Increased Anxiety and Impaired Cognitive Function in C9ORF72 Mice with 450 Repeats

(A and B) Behavioral performances in WT, C9^{450B}, and C9^{450C} mice at 4, 12, and 18 months of age (n = 25 mice per group at 4 months and n = 16 [WT], n = 14 [C9^{450B}], and n = 10 [C9^{450C}] at 12 and 18 months of age). (A) Spatial learning and memory performance on a Barnes maze showing the number of errors in finding the escape chamber at days 1–3, 4–6, and 7–9. (B) Working memory performance on a radial maze showing errors per trial over 10 days of testing. (C and D) Anxiety-related behaviors in WT, C9^{450B}, and C9^{450C} male mice at 4, 12, and 18 months of age (n = 11 [WT] and n = 13 [C9^{450B}] at 4 months, and n = 9 [WT], n = 4 [C9^{450B}], and n = 7 [C9^{450C}] at 12 and 18 months of age). (C) Anxiety-related behavior determined by marble burying test showing the percent of marbles buried during a 20 min trial, and (D) elevated plus maze showing the percent of time spent on the open arm. (E and F) (E) Representative images and (F) quantification of DAPI-positive nuclei in the hippocampal dentate gyrus and CA1 region in 4- and 12-month-old WT, C9^{450B}, and C9^{450C} mice (n = 4–5 per group). Error bars represent SEM. n.s., not significant, *p < 0.05 and **p < 0.01 using one-way ANOVA.

that in our transgenic mice, poly(GP) proteins are mostly translated from sense strand RNAs.

Selective reduction of repeat-containing RNAs with attenuated RNA foci and RAN translation was confirmed by single-

dose injection of two additional ASOs (ASO3 and ASO4) targeting sequences 5' to the expansion within C9ORF72 intron 1 (Figures 7B and S8A). Two weeks after treatment, repeat-containing C9ORF72 RNAs in cortex and spinal cord were reduced

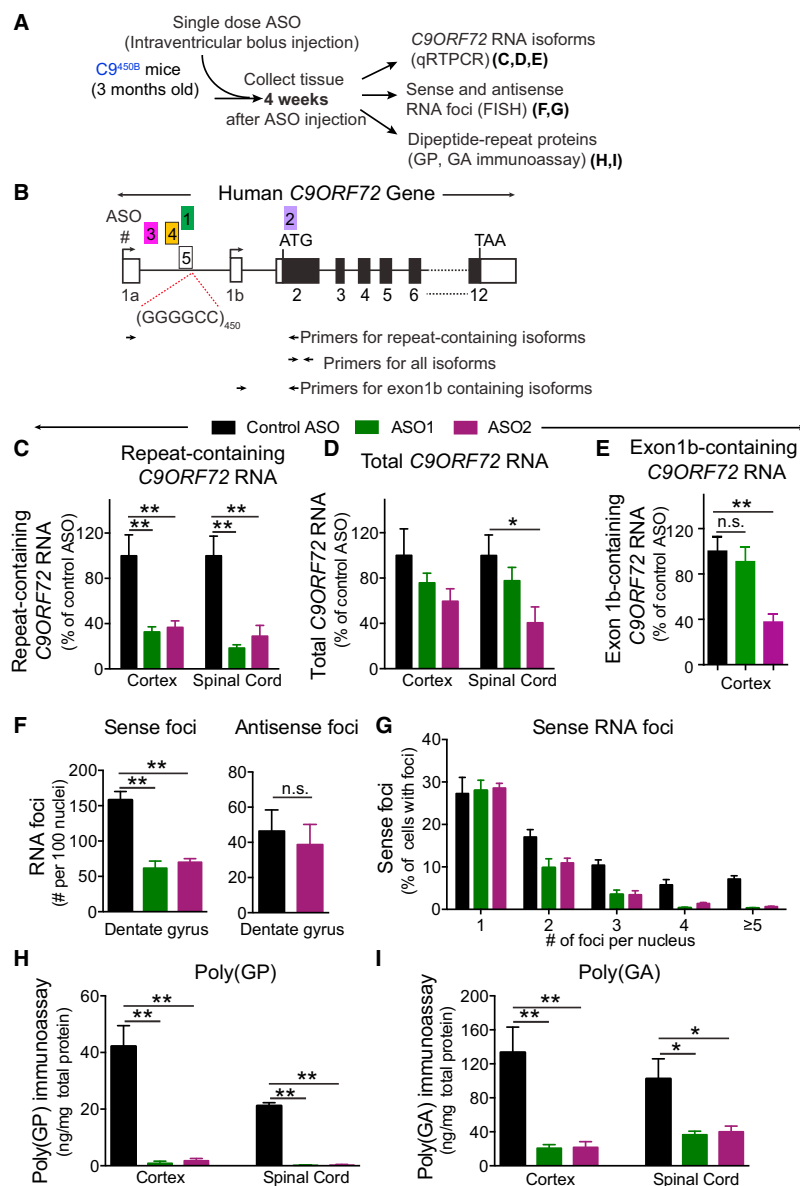


Figure 7. Sustained Reduction in RNA Foci and DPR Proteins from a Single Dose of ASOs Targeting *C9ORF72* Repeat-Containing RNAs

(A) Schematic of injected ASOs targeting the sense strand *C9ORF72* transcripts for degradation in 3-month-old *C9^{450B}* mice.

(B) Schematic of the *C9ORF72* gene showing the GGGGCC repeats within the first intron, the two transcription initiation sites (arrows), the positions of five ASOs, and primers for detection (by qRT-PCR) of various *C9ORF72* RNAs.

(C–E) Expression of (C) repeat-containing, (D) total, and (E) exon 1b-containing *C9ORF72* RNAs determined by qRT-PCR in mice treated either with ASO1 targeting only the repeat-containing RNAs, ASO2 targeting all *C9ORF72* variants, or a control ASO.

(F and G) (F) Number of sense and antisense foci (per 100 nuclei) determined by FISH and (G) quantification of sense foci per nucleus in hippocampal dentate gyrus.

(H and I) Levels of (H) poly(GP) or (I) poly(GA) in the cortex and spinal cord of *C9^{450B}* mice treated with ASO1, ASO2, or a control ASO, as measured by immunoassay. Error bars represent SEM in $n = 5–6$ biological replicates. * $p < 0.05$, ** $p < 0.01$, n.s. not significant using one-way ANOVA.

to 40% of a PBS-treated group, with total *C9ORF72* RNAs remaining at 80% of their initial level (Figures S8B and S8C). Sense foci were reduced by half; antisense foci were unaffected. SDS-soluble poly(GP) levels were reduced by more than 80% in cortex and 50% in spinal cord (Figure S7D), and poly(GP) aggregates, quantified in a blinded fashion, were significantly decreased in ASO3- or ASO4-treated mice (Figures S8E and S8F). Thus, intron 1-targeting ASOs rapidly decrease both soluble and insoluble DPR proteins throughout the mouse CNS.

Finally, 9-month-old *C9^{450B}* mice were injected with an ASO targeted to repeat-containing RNAs (Figure 8A). Repeat-containing *C9ORF72* RNAs in the cortex and spinal cord were sharply

reduced (to 23% and 12% of control levels, respectively) within 2 weeks of treatment (Figure 8B). Consistent with our assays with an independent cohort (Figures 6A–6D), *C9^{450B}* mice injected with a control ASO developed increased anxiety (in marble burying test and elevated plus maze) and impaired cognition (in Barnes and radial mazes) (Figures 8C–8F). Single-dose ASO injection at 9 months alleviated these age-dependent deficits (Figures 8C–8F). In fact, at 15 months (6 months after injection), the beneficial effects were sustained, with a trend suggestive of further improvement. Correspondingly, SDS-soluble poly(GP) and poly(GA) levels remained lower in 16-month-old mice treated with the repeat-targeting ASO (Figure 8H), even though repeat-containing *C9ORF72* RNA levels had recovered to their initial level (Figure 8G).

DISCUSSION

A key question regarding pathogenic mechanisms in *C9ALS/FTD* has been whether the repeat expansion causes disease through loss of *C9ORF72* function, gain of toxicity from repeat RNA, or both. By producing and analyzing mice with a chronic reduction of *C9ORF72* or that express the human *C9ORF72* gene with different sizes of expanded repeats, we have identified gain of toxicity as a central disease mechanism in a mammalian

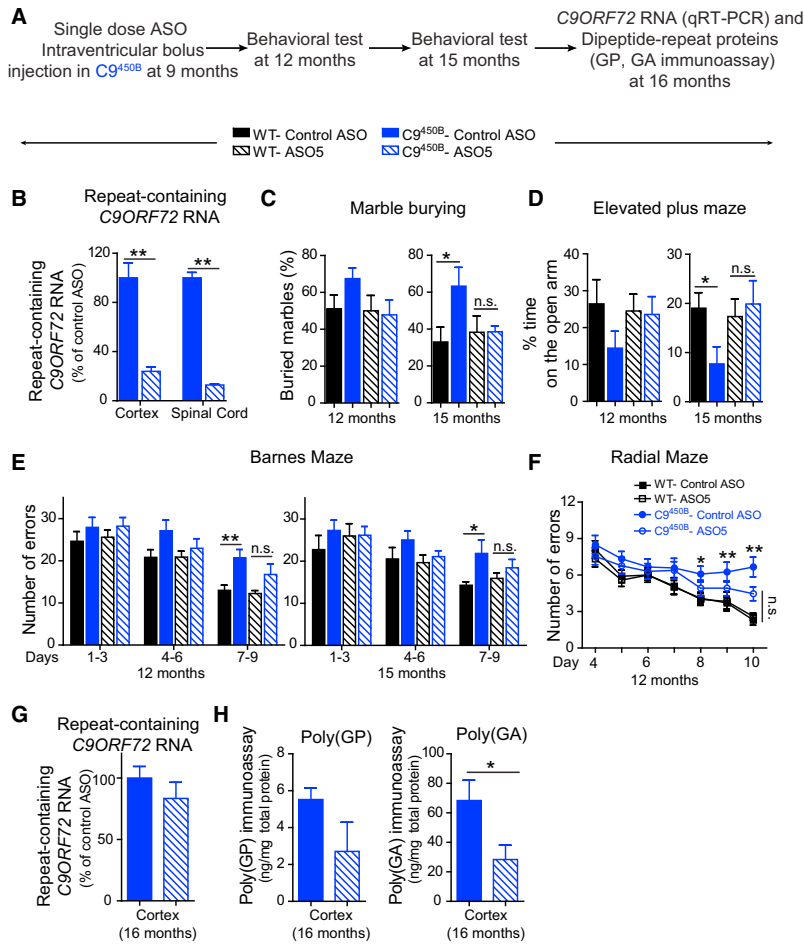


Figure 8. Single-Dose ASO Treatment Alleviates Age-Dependent Behavioral Deficits in *C9ORF72* Mice Expressing 450 Repeats

(A) Schematic of the experimental procedure for a single-dose ASO targeting degradation of the sense strand *C9ORF72* RNA in 9-month-old *C9^{450B}* mice.

(B) Expression of repeat-containing *C9ORF72* RNAs was determined by qRT-PCR 2 weeks after injection of ASO5 or control ASO ($n = 6$ per group). (C and D) Anxiety-related behaviors determined by (C) marble burying test and (D) elevated plus maze in 12- and 15-month-old WT and *C9^{450B}* mice treated with either control ASO or ASO5 ($n = 5-7$ per group).

(E and F) Cognition-related behaviors determined on (E) a Barnes maze and (F) a radial maze in 12- and 15-month-old WT and *C9^{450B}* mice treated with either control ASO or ASO5 ($n = 10-13$ per group).

(G and H) (G) Expression of repeat-containing *C9ORF72* RNAs and (H) levels of poly(GP) or poly(GA) proteins in the cortex of 16-month-old *C9^{450B}* mice treated at 9 months with ASO5 or control ASO. Error bars represent SEM in biological replicates * $p < 0.05$, ** $p < 0.01$, n.s. not significant, using Student's *t* test.

By disrupting both *C9orf72* alleles, we determined that complete elimination of *C9ORF72* from mice produces splenomegaly, enlarged cervical lymph nodes, and premature death. While loss of *C9ORF72* during embryonic development has been reported to produce motor deficits in zebrafish (Ciura et al., 2013) and *C. elegans* (Therrien et al., 2013), systemic (our work here and

Atanasio et al., 2016; O'Rourke et al., 2016) or nestin-cre (Koppers et al., 2015)-mediated ablation of *C9orf72* in mice is not associated with motor degeneration, defects in motor function, or altered survival. In addition, no loss-of-function mutations in *C9ORF72*, including nonsense or frameshift mutations, have been linked to ALS or FTD (Harms et al., 2013), and reduced expression of expanded RNAs following hypermethylation of the *C9ORF72* promoter may actually be neuroprotective (Liu et al., 2014; McMillan et al., 2015; Russ et al., 2015). While our studies reinforce conclusions that *C9ORF72* plays an important role in immune cells (Atanasio et al., 2016; O'Rourke et al., 2016), chronic reduction of *C9ORF72* alone is not sufficient to cause ALS/FTD symptoms in mice. A key remaining question, now testable with the mice we report here, is whether a reduction in *C9ORF72* synergizes with repeat-mediated gain of toxicity to potentiate disease.

Our analysis uncovered an unexpected association between hexanucleotide repeat length and accumulation of RNA foci and DPR proteins. When expressed at seven times the level of an endogenous *C9orf72* allele, no detectable DPR proteins accumulate in mice expressing 110 repeats (Figure 4F). Conversely,

nervous system. Mice expressing RNAs with 450 repeats from two independent lines developed age-dependent, anxiety-like behavior and impaired cognitive function by 12 months of age, accompanied by mild hippocampal neuronal loss. In contrast, mice hemizygous for *C9ORF72* that express 50% of *C9orf72* mRNA, as observed in C9ALS/FTD patients (DeJesus-Hernandez et al., 2011; Gijselink et al., 2012; van Blitterswijk et al., 2015), did not develop an ALS/FTD-like phenotype or any other phenotype in the CNS or periphery. Hence, by a systematic comparison of multiple mouse lines, we now demonstrate that reduction of *C9ORF72* expression is not sufficient to trigger neurodegeneration in mice, while pathological hallmarks of C9ALS/FTD, neuronal loss and cognitive dysfunction develop in mice expressing expanded repeat RNA.

The function of the 54 kD *C9ORF72* protein has not been established, but its sequence homology to the DENN protein family predicts a possible role in regulating membrane trafficking (Farg et al., 2014; Levine et al., 2013; Zhang et al., 2012). The *C9orf72* gene is expressed not only in the CNS but also in peripheral tissues, including spleen (Suzuki et al., 2013), bone marrow, lymphocytes, and macrophages (BioGPS data; <http://biogps.org/>).

mice with lower RNA levels but with 450 repeats produce both soluble DPR proteins (Figures 4F–4H) and insoluble aggregates (Figures 4I–4K). That no DPR proteins were detected in C9¹¹⁰ mice could be due to (1) RAN translation initiation of short repeats being less efficient than of long repeats; (2) shorter DPR proteins having shorter half-lives than longer DPR proteins; (3) shorter repeat-containing RNAs being less effectively transported to the cytoplasm, the presumed site of RAN translation; or (4) a combination of options 1–3.

While a prior study demonstrated that robust adeno-associated-virus-mediated expression of 66 GGGGCC repeats led to the aggregation of DPR proteins throughout the murine CNS (Chew et al., 2015), we demonstrate here that expression of a short repeat does not generate such pathology in mouse brain when expressed at seven times the levels of RNA from a single *C9orf72* allele. Not only do long and short repeats have differing capacities to generate DPR proteins, long and short DPR proteins have different intracellular localization and aggregation profiles. For instance, DPR proteins translated from 66 repeats produced a level of accumulated poly(GP) that reached 1.8% of total brain protein by 6 months of age and formed primarily nuclear aggregates (Chew et al., 2015), whereas we and others (O'Rourke et al., 2015; Peters et al., 2015) have shown that in mice with 400–500 repeats, DPR proteins form cytoplasmic inclusions similar to those observed in C9ALS/FTD patients. Short DPR proteins are likely to be more soluble and sufficiently small to freely diffuse through nuclear pores, thereby facilitating the intranuclear aggregation of DPR proteins seen in mice expressing 66 repeats (Chew et al., 2015). Similarly, the nucleolar disruption and acute toxicity resulting from exposing cultured cells to 10 μ M of 20-mer poly(PR) or poly(GR) proteins (Kwon et al., 2014) differ markedly from the primarily cytoplasmic aggregates observed in mice expressing 450 repeats and C9ALS/FTD patients (Ash et al., 2013; Gendron et al., 2013; Mackenzie et al., 2015; Mori et al., 2013a; Zu et al., 2013).

While the minimal pathogenic repeat size in *C9ORF72* patients is not established, somatic expansion can take the germline-transmitted repeat to 3,000–5,000 repeats in the most affected brain regions (Gijssels et al., 2015; van Blitterswijk et al., 2013). We have found minimal somatic expansion in either the CNS or peripheral tissues of mice. Also differing from C9ALS/FTD patients, our C9⁴⁵⁰ mice did not display TDP-43 mislocalization or aggregation, although increased levels of phosphorylated TDP-43 were detected. In addition, we did not observe mislocalization of proteins involved in nucleocytoplasmic transport. Lack of these features may explain the relatively mild phenotype observed in our transgenic mice despite expression of up to 24 times a single endogenous *C9orf72* allele. While abundant RNA foci did not increase with age in our mice, an age- and repeat-length-dependent increase in the number and size of cytoplasmic DPR aggregates was seen, with soluble poly(GP) levels decreasing with age (Figure 4H). In C9ALS/FTD, the anatomical distribution of DPR protein pathology does not correlate with neurodegeneration (Schludi et al., 2015). These findings suggest that certain neuroanatomical regions are less susceptible to DPR proteins, or that soluble DPR proteins are toxic and DPR protein aggregates are neuroprotective. In support of the latter, poly(GA) can recruit poly(GR) into inclusions and partially

decrease poly(GR) toxicity in *Drosophila* and cultured cell models (Yang et al., 2015). On the other hand, aggregation of poly(GA) was recently reported to be necessary for its toxicity, which is mediated through sequestration of HR23 and nucleocytoplasmic transport proteins (Zhang et al., 2016).

Despite uncertainty about the contributions of RNA foci or DPR proteins to neurodegeneration, an “on mechanism” therapy would be one that reduces repeat-containing RNAs, thereby attenuating both potential toxicities. We have established the potency of multiple ASOs for selective reduction of repeat-containing *C9ORF72* RNAs in the mammalian CNS while having minimal effect on total *C9ORF72* RNAs. Importantly, single-dose ASO (1) significantly mitigated the accumulation of sense RNA foci (without increasing antisense foci) and of poly(GP) and poly(GA) proteins and (2) significantly attenuated the development of behavioral deficits even 6 months after treatment. Thus, our studies establish a repeat-dependent gain of toxicity as a crucial pathological mechanism of C9ALS/FTD and, most importantly, validate the feasibility of employing ASO therapy to mitigate the toxicity from repeat RNAs without exacerbating a potential loss of C9ORF72 function.

EXPERIMENTAL PROCEDURES

Please see the [Supplemental Experimental Procedures](#) for more information

Generation of *C9ORF72* BAC Transgenic Mice

The human BAC construct expressing a truncated *C9ORF72* gene with 450 repeat expansions was obtained from BACPAC resource center at Children's Hospital Oakland Research Institute (clone ID: CH523-111K12) and was injected into the pronuclei of fertilized C57BL/6J hybrid eggs and implanted into pseudo-pregnant female mice. Mice used in this report were then backcrossed to C57BL/6 for a minimum of three generations. All experimental procedures were approved by the Institutional Animal Care and Use Committee of the University of California, San Diego.

RNA Extraction and qRT-PCR

Procedures are detailed in the [Supplemental Information](#). Primers and probe sequences are listed in [Table S1](#).

Generation of Antibodies Recognizing DPR Proteins

Peptide antigens (C-Ahx-(GA)₈-amide, C-Ahx-(GR)₈-amide, C-Ahx-(PR)₈-amide, C-Ahx-(GP)₈-amide, and C-Ahx-(PA)₈-amide) were used to immunize rabbits to generate antibodies against DPR proteins. Pre-immune serum from each rabbit was tested using peptide antigens and tissue from C9ALS/FTD cases by immunoblot and immunohistochemistry, respectively, and confirmed negative. Antiserum or affinity-purified antibodies were used. Antibodies specific for each DPR protein are herein annotated Rb4333 poly(GA), Rb4335 poly(GP), Rb4995 poly(GR), Rb15986 poly(PA), and Rb15986 poly(PR).

Immunofluorescence Staining and X-gal Staining

Sections from paraformaldehyde-fixed tissues were stained using standard protocols with antibodies against GFAP (Chemicon, 1:1000), IBA1 (Wako, 1:500), ChAT (Millipore, 1:300), NeuN (GeneTex, 1:1000), CTIP2 (Abcam, 1:500), poly(GA) (Rb4333, 1:1000), poly(GP) (Rb4335, 1:1000), poly(GR) (Rb4995, 1:1000), poly(PR) (Rb15986, 1:100), poly(PA) (Rb15989, 1:100), TDP-43 (Proteintech, 1:500), RanGAP1 (Santa Cruz, 1:500), Lamin B (Santa Cruz, 1:20,000), and P62 (Abnova, 1:100). Confocal images were acquired on a Nikon Eclipse laser scanning confocal microscope using the Nikon EZ-C1 software. LacZ activity was assessed with X-gal staining solution (1.0 mg/mL of X-gal, 5 mM potassium ferrocyanide, and 2 mM MgCl₂) for 12 hr at 37°C. The sections were examined and photographed with a Nanozoomer.

Injection of ASO in the Mouse CNS

Intra-cerebroventricular (ICV) stereotactic injections of 10 μ L of ASO solution, corresponding to a total of 350 μ g of ASOs (Ionis Pharmaceuticals), were administered into the right ventricle using the following coordinates: 0.2 mm posterior and 1.0 mm lateral to the right from the bregma and 3 mm deep. Mice were treated either with PBS; a control ASO; ASOs targeting total *C9ORF72* (ASO2); or repeat-containing *C9ORF72* variants (ASO1, ASO3, ASO4, or ASO5). The sequences of the ASOs are available in Table S2.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, eight figures, and two tables and can be found with this article online at <http://dx.doi.org/10.1016/j.neuron.2016.04.006>.

AUTHOR CONTRIBUTIONS

J.J., Q.Z., T.F.G., P.J., S. Sabri, S.-C.L., M.M., C.J.H., P.V.D., D.T.U., A.D., S.M.H., L.P., C.F.B., S.D.C., J.R., F.R., D.W.C., and C.L.-T. designed the experiments and analyzed the data. J.J., Q.Z., T.F.G., S. Sabri, J.E.S., P.J., K.D., D.S., S.C., A.S., S. Sun, S.-C.L., M.M.-D., B.M., J.E., M.K., M.B., O.P., A.W., C.J.H., L.D.M., L.M.D., A.D., and D.T.U. performed the experiments. M.C.A., D.S., D.A.S., L.T., C.J.J., P.J.D.J., D.E., S.M.H., and C.E.S. contributed key reagents and methodology. J.J., Q.Z., T.F.G., D.W.C., and C.L.-T. wrote the manuscript.

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